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SYNTHESIS OF SILYL PEROXIDE OF COELENTERAZINE (*OPLOPHORUS* **LUCIFERIN) ANALOGUE FOR PRECURSOR OF LUMINESCENCE**

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Abstract : Unstable tert-butyldimethylsilyl peroxide of coelenterazine (Oplophorus luciferin) analogue has *been synthesized b_v radical reaction of rert-hutyldimethylsilyl hydroperoxide. This compound may be a* key intermediate model in the bioluminescence and chemiluminescence of coelenterazine.

Aequorin **(1)** is a calcium-binding protein found in jellyfish, *Aequorea vicutoria,* to emit blue light (46Onm) by the action of calcium ions to this luminescence system. 1 Molecular mechanism studies of the luminescence reaction, and applications of the apoprotein 2 and chromophore 3 have recently progressed. Aequorin is known to have a chromophore, coelenterazine (*Optophorus* luciferin) 2, as the light emitting species linking to this protein through a peroxidic bond as illustrated in 1. In 1978, Shimomura and Johnson reported that a yellow compound obtained by reduction of aequorin with NaHSO₃ has a tertiary alcohol at the imidazolone carbon to which the p-hydroxybenzyl group is attached.⁴ Kishi and Shimomura et al. supported this result by measuring ¹³C-NMR spectrum of 1 and concluded its structure through incorporation experiment of ${}^{18}O_2$.⁵ Recently we reported that the structure of yellow compound, which is **diktly connected** to the original structural information of the aequorin chromophore, **is** the 5oxy structure 3.6

Coelenterazine and its analogues emit blue light in organic solvents , **for example dimethyl sulfoxide or dimethylformamide (DMF), under aerobic condition without apoprotein.7 While molecular mechanism of this chemiluminescence has been studied for many years, it is still uncertain.8**

We has suggested that hydroperoxide 4 of coelenterazine is involved in these luminescence reactions **as illustrated in Scheme I. Our interest in the chemistry of coelenterazine peroxide has prompted us to synthesize peroxides related to 4 in an effort to evaluate our proposal.**

Scheme 1. Posturated mechanism of luminescence reaction.

A number of synthetic approaches to peroxides related to 4 have been examined (illustrated in Scheme 21. These approaches all began wiih the coelenterazine analogue 7, having ter?-butyl group at 2-position of the imidazopyrazinone, which was readily prepared by coupling ⁹ between the 2-aminopyrazine 5^{10} and the tert-butyl glyoxal 6¹¹ in 90% yield as shown in Scheme 2. In the initial approach, hydrogen peroxide **cuprous chloride system and photooxygenation system were employed as the oxidizing system, however not the hydroperoxide 8 but 2-amidpyrazine 9 was formed under these conditions. Subsequently it was** found that tert-butyl peroxide compound 10¹² could be particularly easy obtained in 30 \sim 90% yield¹³ **using anhydrous tert-butyl hydroperoxide - cuprous chloride system 14. Similarly, terf-butyldimethylsilyl** hydroperoxide (TBDMSOOH) 11¹⁵- cuprous chloride system gave *tert* -butyldimethylsilyl peroxide 12.

The experimental procedure is as follows; to a distilled methylene chloride solution of coelenterazine analogue 7 under an argon atmosphere was added TBDMSOOH (2.0 equiv) at -20^oC followed by addition of a catalytic amount of cuprous chloride. The resulting mixture was stirred for $3 \sim 30$ hr at -15 °C, then poured into a mixture of ice-cooled, 0.01N Na₂S₂O₃ aqueous solution and methylene chloride, and **extracted with methylene chloride. The combined methylene chloride extracts were thoroughly washed with** distilled water, dried over anhydrous $Na₂SO₄$ and evaporated under reduced pressure at $5 \sim 10^{\circ}C$. The **residue was chromatographed thrice on silica gel column by using I** : **30 mixture of ethyl acetate and n**hexane. methylene chloride and 1 : 20 mixture of ethyl acetate and n-hexane at -50^oC ~ -40^oC respectively **to give tert-butyldimethylsilyl peroxide 12 in 2% yield. About 30% loss of 12 occurred in each chromatography due to instability of this compound. Product structure was assigned on the basis of spectroscopic data 16 and confirmed by chemical transformations. For example, 12 was reduced to 13** with dimethyl sulfide, which was identified with authentic sample ¹⁷, reduced to 7 with Na₂S₂O₄ and converted directly to 14¹⁸ by treatment of 12 for a few minutes at 0^oC with silica gel. While other silyl **hydroperoxides. for example, triphenylsilyl hydroperoxide, and other radical sources were employed, never desired silyl peroxide compound was obtained.**

Scheme 2. Synthesis of peroxides of coelenterazine **analogue.** Reagents; (a) HCOCOC(CH₃)₃ (6), HCl, H₂O, 1,4-Dioxane, 100^oC; (b) (CH₃)₃COOH, CuCl, CH₂Cl₂, -15^oC ; (c) [O] ; (d) TBDMSOOH (11), CuCl, CH₂Cl₂, -15^oC ; (e) (CH_3) , S, THF, $0^{\circ}C$; (f) $Na_2S_2O_4$, CH₃OH, H₂O, $0^{\circ}C$; (g) silica gel, $0^{\circ}C$; (h) DMF, 20° C or n -Bu₄NF, AcOH, THF, -78 $^{\circ}$ C \sim 20 $^{\circ}$ C.

Purified 12 is very unstable. The stability in solution decreases in the order : chloroform, methanol, DMF. 12 can be kept for a day at 20^oC in chloroform, in which 12 is most stable, but it rapidly decomposes in methanol to give compound 14 and a few compounds. In DMF under either aerobic or anaerobic condition, 12 emitted weak light for 2 days.¹⁹ Desilylation of 12 in tetrahydrofuran at -78°C with n-Bu₄NF in the presence of acetic acid should lead to the hydroperoxide or the hydroperoxide anion and these species fragmented to emit blue light (490 nm) at -30°C~-20°C under anaerobic condition.¹⁹ Unfortunately, the hydroperoxide 8 or the hydroperoxide anion could not be detected as an intermediate. Thus, 8 or the anion would appear to be very labile in this desilylation condition. Further work to detect or isolate the hydroperoxyde 8 is now in progress.

In summary, this work has first demonstrated that peroxy coelenterazine analogue for the luminous precursor is formed by using TBDMSOOH - cuprous **chloride system. Further,** it **has** been shown that silyl peroxide of coelentelazine leads to precursor of luminous species. This work **would provide further mechanism on the bioluminescence and chemiluminescence of coelenterazine.**

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References and Notes

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- 12. 10 ¹H-NMR (CDCl₃, 20^oC, 270MHz), δ 0.92(9H, s), 1.44(9H, s), 3.87(3H, s), 4.09(1H, d, J= 14.2 Hz), 4.21(1H, d, J= 14.2 Hz), $6.67(1H, s)$, $6.94(2H, d, J= 8.9 Hz)$, $7.15\sim7.30(3H, m)$, 7.40(2H, d, J= 6.9 Hz), 8.06(2H, d, J= 8.9 Hz). UV λ max. (MeOH) 432nm(ε = 19700), 303nm(ε = 6670), 248nm(ε = 6410). MS m/z 476 (M+1]⁺, mp 134~135°C (decompose).
- 13. The yield of this reaction is ruled by lots of anhydrous tert-butyl hydroperoxide solution.
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- 15. To a dried ether solution of anhydrous H_2O_2 (5.0 equiv)²⁰ was added tert-butyldimethylsilyl chloride (1.0 equiv) and imidazole (1.0 equiv) at 0° C followed by extractive isolation with n-hexane and evaporation under reduced pressure at 15^oC. The residue was chromatographed on silica gel at 0^oC by using methylene chloride to give tert-butyldimethylsilyl hydroperoxide (88% yield, mp 44~45°C).
- 16. **12** ¹H-NMR (CDCl₃, 20^oC, 270MHz), δ -0.12(3H, s), -0.11(3H, s), 0.75(9H, s), 1.44(9H, s), 3.86(3H, s), 4.20(1H, d, J= 13.9Hz), 4.26(1H, d, J= 13.9Hz), 6.70(1H, s), 6.90(2H, d, J= 8.9Hz), 7.2~7.4(3H, m), 7.45(2H, d, J= 6.9Hz), 8.03(2H, d, J= 8.9Hz). UV λ max. (MeOH) 43lnm(ε = 23 IOO), **3oihIII(E= 'X50), 247nm(s= 8180).** MS **m/z 534** [M+I]+. **mp** 107-108oC (decompose).
- 17. Authentic sample of 13 was prepared as shown in the previous report ⁶. 13 ¹H-NMR (CDCl₃, 20°C, 270MHz). 6 1.44(9H. s), 3.86(3H, s). 4.07(lH, d, J= 15Hz). 4.17(lH, d, J= 15Hz). 6.62(1 H, s). 6.94(2H, d, J= 9.OHz), 7.1-7.2(3H, m), 7.22(28, d, J= 6OHz), 8.06(2H, d, J= 9.OHz).
- 18. 14 'H-NMR(CDC13, 2OoC, 270MHz), & 1.46(9H, s), 3.87(3H, s), 4.21(2H, s), 6.94(2H, d, J= 9.2Hz). 7.15-7.40(3H, m), 7.43(2H, d, J= 7.OHz), 8.38(2H, d, J= 9.2Hz). **UV hmax. (MeOH)** 472nm (ε = 1800), 362nm (ε = 21000), 253nm (ε = 15900). MS m/z 402 [M+1]⁺. mp 156~157°C.
- 19. The chemiluminescence of **12** will be described in a subsequent paper.
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